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PHYTOCHEMICAL INVESTIGATION AND ANTIBACTERIAL ACTIVITY TEST OF THE ROOT EXTRACT OF ALOE CALIDOPHILA

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ABSTRACT

Aloe calidophila is one of the endemic Aloe species traditionally used for the treatment of various illnesses including stomach pain, malaria, hypertension, diarrhea and wound healing in Sidamo floristic region and southern part of Ethiopia. The aim of this study was investigate chemical constituents and subsequently characterize isolated compounds from root extract of Aloe calidophila using FTIR, ¹HNMR and ¹³CNMR spectroscopic techniques as well as testing its antibacterial activity. The phytochemical screening tests of the dichloromethane/methanol (1:1 v/v) and methanol root extracts revealed the presence of alkaloids, anthraquinones, flavonoids, phenols, terpenoids, steroids and saponins. The chromatographic separation of dichloromethane/methanol (1:1 v/v) crude extract resulted in three compounds (ABM1, ABM2 and ABM3) using n-hexane/ethyl acetate solvent system. The chemical structures of the compounds were found to be aloes aponarin I, oleic acid and β -sitosterol, respectively, based on spectroscopic (FTIR and NMR) data and literature reports. The crude extracts and isolated compounds were evaluated for their antibacterial activities against four bacterial strains specifically (E. coli ATCC25922, S. aureus ATCC25923, P. aeruginosa ATCC27853 and S. pyogenes ATCC19615). Disc diffusion method was used for the test. The antibacterial activity tests revealed that the isolated compounds to show relatively better activities than the crude extracts. The isolated compounds showed comparative activities when compared to each other with compound AMB3 to be slightly highest activity against the bacterial strains used in the study. The present finding indicated that the roots Aloe calidophila could be good sources of compounds to be used as leads in the discovery of antibacterial agents provided that in vitro tests are carried out in several bacterial strains.

KEYWORDS

Aloe calidophila, Aloesaponarin I, Oleic acid, β-sitosterol, Antibacterial activity and Phytochemical screening.

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INTRODUCTION

The plant species that belong to the genus *Aloe* are the oldest medicinal plants known to human¹. They are native to sub-Saharan Africa, many islands of western Indian Ocean, including Madagascar and Saudi Arabian Peninsula^{2,3}. Reports revealed the use this plant species for treatment of several

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human illnesses stomach such as ailments. gastrointestinal problems, skin diseases. constipation for radiation injury, for its antiinflammatory effect, for wound healing and burns, as an antiulcer and diabetes⁴⁻⁶. There are also literature reports that state its wide use in the preparation of skin care products, cosmetics and as nutraceutical^{7,8,6}. The leaves and roots *Aloe* species are well known as principal sources of many interesting phytochemicals such as anthraquinones, preanthraquinones, alkaloids. anthrones. chromones, glycosides, carbohydrates, coumarins, flavonoids, naphthalene, steroids, volatile oils etc^{9,10}.

In Ethiopia and Eritrea, about 46 species of Aloe have been identified so far with a high proportion of endemics adapted to harsh climates. It has been reported that the leaf latex of several Aloe species and their constituents possess wide spectrum of biological activities such as antimicrobial^{11,12}, antimalarial^{9,13} and antiglycation¹⁴. The aloe species in Ethiopia are distributed in all floristic region of the country including Afar, Arsi, Bale, Gamo Gofa, Gojam, Gonder, Harerge, Kefa, Shewa, Sidamo, Tigray, Wellega and Welo floristic regions (Sebsebe and Nadal, 2010)¹⁵. Accordingly, the majority of them are found in Sidamo floristic region (with 14 species) followed by Harerge, Bale, Shewa, Tigray and Welo floristic regions (with 10, 8, 8, 7 and 7 species respectively). Whereas few species found in Afar and Wellega floristic regions (each possess one Aloe species) and one Aloe species (Aloe macrocarpa) found in many floristic regions including Arsi floristic area (Sebsebe and Nadal, 2001). The distribution of *Aloes* in Ethiopia can be correlated to vegetation types, and showed that the endemic and near-endemic species occurred predominantly in two vegetation types. These are: (a) dry montane evergreen forest (and associated montane evergreen scrub or montane grassland), and (b) Acacia-Commiphora woodland and bushland¹⁶. Some of the *Aloes* that occur in dry montane evergreen forest include Aloe debrana, Aloe adigratana, Aloe percrassa, Aloe pulcherrima, Aloe elegans, Aloe camperi and Aloe yavellana.

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Acacia-Commiphora wood land and bush land also host *Aloe* species such as *Aloe* calidophila, *Aloe* ellenbeckii, *Aloe* gilbertii, *A. Aloe* friisii, *Aloe* retrospiciens, *Aloe* mcloughlinii, *Aloe* pirottae, *Aloe* otallensis and *Aloe* trichosantha. *Aloe* species such as *Aloe* bertemariae and *Aloe* citrine are reported from desert and semi-desert scrubland and *Aloe* ankoberensis is from Afro-alpine vegetation of Ethiopia¹⁷.

Aloe calidophila (Figure No.1) is one of the endemic species of Ethiopia which grows mostly in Acacia-Commiphora woodland/bushland or open wooded grassland in altitude range of 1280-1620m in Dida Cheena Plains, between Moyale and Mega, in Sidamo floristic region¹⁷. The different parts of Aloe calidophila are used for the treatment of various diseases in traditional or folk medicine in Ethiopia. For instances, its leaf and roots are traditionally used in Sidama region, South nations around Gamo Gofa, and in other central highlands of the country for the treatment of various illnesses such as stomach pain, malaria, hypertension, diarrhea, endocrine system, repel flies from infected eye, wound healing, and anthelmintic¹⁸. Despite its wide use in traditional medicine, there are no exhaustive studies. In other words, there are only a limited study reports on this species. For instance, a study on phytochemical and biological activities of latex from its leaves revealed the presence of phytochemicals that have been presumed to be responsible for its antileishmanial activity. Some of them are 4-dihydroisocoumaringlucoside, 7-aloin B, 8-aloin A, 9-aloinoside B, 10-aloinoside A, 11microdontin B, 12-microdontin A^{19,20}. Moreover, compounds isolated from the latex have been reported to show a better anti-promastigote activity than some standard antileishmanial drugs such as Amphotericin B^{21} . To the knowledge of our team, there are no reports on phytochemical investigations and isolation of compounds from the root parts of Aloe calidophila. Therefore, the aim of the present study was to carry out phytochemical screening on root extract of Aloe calidophila and its chemical composition, and also evaluation of antibacterial

activities of the crude extracts and compounds isolated from crude extracts.

MATERIAL AND METHODS Collection of plant material

The root of *A. calidophila* was collected from Dida Cheena, an area between the towns of Yabello and Mega in the Southern Nation, Nationalities and Peoples' Region of Ethiopia. The area is about 550 km south of Addis Ababa and 221km south of Hawassa town. The plant material was authenticated by a botanist Professor Sebsebe Demissew, Department of Biology, Addis Ababa University.

Preparation of plant material

The collected plant material (root part) was washed with tap water and chopped to small pieces and airdried at room temperature for one month. The dried plant material was ground using a grinding machine to facilitate the extraction process.

Extraction

500g of the powdered root of *A. calidophila* was soaked in 4 L of dichloromethane/methanol (1:1 v/v) at room temperature in Erlenmeyer flask. After shaking the contents well, the flask containing the solution was placed on an orbital shaker and agitated at the speed of 120 revolutions per minute for 72 hours. Then the solution was filtered using Whatman filter paper. The filtered solution was concentrated using rotary evaporator (Heidolph, UK) at temperature of 40°C. The marc left was further extracted with 100% methanol following similar steps used above (Scheme No.1).

Phytochemical screening tests

Phytochemical screening tests or identification of secondary metabolites were performed on the crude extract of dichloromethane/methanol (1:1 v/v) and methanol extract using standard procedures to investigate the presence of some secondary metabolites such as alkaloids, anthraquinones, flavonoids, glycosides, phenols, saponins, steroids, tannins and terpenoids using preliminary phytochemical analysis²¹.

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Isolation and characterization of compounds

Nine grams (9g) of dichloromethane/methanol (1:1 v/v) dried crude extract was adsorbed onto 18g of silica gel and subjected to column chromatographic isolation (180g silica gel 60-120 mesh size used as the stationary phase). The column was eluted with increasing gradient of n-hexane in ethyl acetate solvent system. The elution process was started by 100% n-hexane and progressed through various ratios of an increasing portion of ethyl acetate in nhexane. A total of 100 fractions were collected and each fraction was checked by TLC. The spots on the TLC plates were visualized using UV light (254 and 365nm), (UV-Tcc chamber). The collected fractions were concentrated using rotary evaporator. Fractions were tested using TLC and those with the same TLC profiles were combined. The column chromatographic separation led to isolation of three compounds. The fraction number 93-95 gave compound ABM1 (65:35); fraction number 63-74 gave compound ABM2 (80:20) and the fraction number 40-45 gave compound ABM3 (85:15). Then, the structural elucidations of the compounds were carried out based on data obtained from spectroscopic data such FTIR (Perkin Elmer BX infrared spectrometer) and NMR (Bruker Avance 400 MHz) data. The NMR spectroscopic analyses were carried out at The Department of Chemistry, Addis Ababa University and the FTIR analyses were done at The Department of Chemistry, Hawassa University, Ethiopia.

RESULTS AND DISCUSSION

The yield of extracts, qualitative phytochemical tests, characterization of isolated compounds, and antibacterial activity tests are discussed in the following sections.

Masses of crude extracts

The resulting extract of Aloe calidophila root was (2.9%) 14.5g (0.97%)and 4.7g for dichloromethane/methanol (1:1 v/v) and methanol, respectively (Table No.1). This data showed that, the crude extract obtained from dichloromethane/methanol (1:1 v/v) was greater than methanol extracts. This indicated that most of

the components the extracts could be medium polar. This finding was consistent with literature reports²². The mass and TLC profile of the dichloromethane/methanol (1:1 v/v) extract were taken as criteria for selecting this extract for column chromatographic separation.

Phytochemical screening

Phytochemical screening of the dichloromethane/methanol (1:1) and methanol root extracts revealed the presence alkaloids. anthraquinones, flavonoids, phenols, terpenoids, steroids and saponins (Table No.1). According to above phytochemical observation, Aloe calidophila is one of the plants rich in many secondary metabolites which could be attributed to be responsible for the traditional medicinal uses and also the plant as such studies may lead to drug discovery and development.

Isolation and characterization of compounds

Three compounds (ABM1, ABM2 and ABM3) were isolated in the present study. The structures of the compounds were determined based on spectroscopic data (IR and NMR) and other physical data such as the melting point. The collected data were analyzed and compared with similar data reported in literature.

Compound ABM1 was isolated as yellow crystal (19.1mg) with a melting point of 190-193°C and an R_f value of 0.4 (35% ethyl acetate in n-hexane). The FTIR spectrum (Appendix 1) of the compound showed a broad band at 3345cm⁻¹ and band at 1697cm⁻¹ indicated the presence of OH and stretching of unsaturated carbonyl group, respectively. Intense bands at 2925 and 2856cm⁻¹ could be attributed to C-H stretching and vibration of C-H bonds of saturated groups. The absorption band around 1460 and 1579cm⁻¹ could be attributed to aromatic ring C=C stretching as well as chelated -COO group²⁴. The ¹HNMR spectrum (400 MHz, DMSO) (Appendix 2) showed the presence of two broad singlets at 2.75ppm and 3.88ppm which correspond to protons of CH₃ and OCH₃ attached to an aromatic $ring^{23}$. There are four sets of aromatic protons of which three of them $(\delta 7.30 \text{ (dd, } 8.3, 1.3 \text{)})$ Hz), $\delta7.61$ (d, J=1.3 Hz), $\delta7.70$ (t, J=7.9 Hz))

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belong to ring A having ABX multiplicity pattern with ortho-para and one meta coupling systems and a singlet peak at $\delta 7.72$. Furthermore, highly deshielded signals at $\delta 12.78$ and $\delta 11.78$ could be attributed to a hydroxyl group peri to carbonyl carbon of aromatic compounds (Mudin et al., 2018). These data suggest that compound is most likely anthraquinone type. The ¹³CNMR (101 MHz, DMSO) spectrum (Appendix 3) revealed seventeen carbon signals, two of which are carbonyl carbons at δ 189.65 and 182.21ppm that are typical of anthraquinone skeleton. The peaks at δ 167.64 and 52.99 reveal the presence of methyl ester group attached to an aromatic ring. The spectrum revealed eleven quaternary carbons. These are six nonoxygenated aromatic quaternary carbons a (at δ122.89, 117.13, 132.77, 124.91, 137.17 and 141.46), two oxygenated aromatic quaternary carbons at (at $\delta 161.89$ and 159.33) and three carbonyl carbons (δ 167.64, 182.21 and 189.65) (Table No.2). The DEPT-135 spectrum (Appendix 4) showed that there are four aromatic methines at δ112.45, 136.57, 118.82 and 124.52. The peaks at $\delta 20.36$ and $\delta 52.99$ could be attributed to a methyl connected to aromatic ring and methyl of ester (a methoxy group), respectively (Table No.2). Thus, based on the above spectral data and comparison with literature reported data, the structure of compound ABM1 was proposed to be similar with 6-dihydroxy-8-methyl-anthracene-7-methyl 1. ester-9, 10-dione or Aloesaponarin I (Figure No.2) (Mudin et al, 2018, Abdissa et al, 2017, Tamiru and Ebsa, 2022)²³⁻²⁵. It was reported that this compound is found in many aloe species such as Aloe gilbertii. Aloe secundiflora, Aloe pulcherrima, Aloe turkanensis and Aloe eleganis^{23,24}. This is the first report of isolation from A. calidophila. The ¹H-NMR, ¹³C-NMR and DEPT-135 spectra data of compound ABM1 are given below (Table No.2).

Compound ABM2

Compound ABM2 (17.3mg) was isolated as yellow crystal with a melting point of $13-14^{\circ}$ C and an R_f value of 0.58 (20% ethyl acetate in n-hexane). In the FTIR spectrum of compound ABM2 (Appendix 5), the bands at 2920cm⁻¹ and 2850cm⁻¹ indicated

the C-H stretch of olefinic group and alkyl groups, respectively. The band at 1707 cm^{-1} indicated C=O stretch of carbonyl groups. This band could be attributed to C=O stretch of carboxylic acid whereas the intense absorption band at 1272cm⁻¹ indicates C-O stretching. The absorption bands at 1467 and 1292cm⁻¹ indicate characteristic bending of C-H bending of CH₂ and CH₃, respectively. Thus, the observed carbonyl group (C=O) and C-O stretching bands in the FTIR spectrum (Appendix 6) suggest that the compound ABM2 is most likely a carboxylic acid. The ¹H NMR (400MHz, CDCl₃) spectrum (Appendix 6) showed the peak at δ 0.9 could indicate the presence of terminal methyl (-CH₃) groups whereas the peaks at $\delta 1.33$ and 1.63could suggest the presence of aliphatic methylene (- CH_2) groups. The peak at $\delta 2.07$ indicated presence of a methylene group that is directly bonded to C=C bond. Instead, the peak at $\delta 2.37$ indicates presence of methylene that is directly bonded to a carboxylic acid group whereas the peak at $\delta 5.38$ indicates presence of olefinic protons in the structure²⁶. The observed spectral data and literature reported data suggest that compound AMB2 could be an aliphatic acid. The ¹³CNMR (101 MHz, CDCl₃) spectrum (Appendix 7) of compound ABM2 showed peak at δ 14.12 could indicate the presence of terminal methyl (-CH₃) group attached to aliphatic acids. The peaks at $\delta 130.03$ and 127.90 the presence of at least one C=C bond whereas a single peak at δ 179.42 indicate C=O stretch of guaternary carbon atom of carboxylic acid. On the other hand, the peaks in the range of $\delta 22.68$ to 33.94 indicated presence of more than one methylene $(-CH_2)$ carbons. The DEPT-135 spectrum (Appendix 8) of this compound also showed single peaks that could indicate the presence of terminal methyl $(-CH_3)$ carbon at $\delta 14.08$, methylene carbons at $\delta 22.71$ to 33.95 and olefinic methine carbons at δ 130.03 and 128.07 (Table No.4). Based on the observed spectral data and comparison of the data from literature reports, compound ABM2 was suggested to be identical with oleic acid (Figure No.3)²⁶⁻²⁸. The ¹³CNMR, ¹HNMR and DEPT-135 spectra data of this compound from A. calidophila root extract

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are reported below (Table No.3). This is the first report of isolation of compound ABM2 from *Aloe calidophila* plant.

Compound ABM3

Compound ABM3 (15.7mg) was isolated as orange crystal with a melting point of 135-137°C and an R_f value of 0.75 (15% ethyl acetate in n-hexane). The FTIR spectrum of compound ABM3 (Appendix 9) showed a broad band at 3474 cm⁻¹ indicating the presence of -OH group in the compound. The absorption bands at 2933 and 2860cm⁻¹ could indicate the presence of C-H stretching for asymmetrical CH₃ and symmetrical CH_2 , respectively. The bands at 1739cm⁻¹ and 1056cm⁻¹ indicated the unsaturation that is C=C and presence of C-O stretches in the compound, respectively. The ¹H NMR (400MHz, CDCl₃) spectrum (Appendix 10) of compound ABM3 revealed the presence of methyl (CH₃), methylene (CH₂), methines (CH) protons, and one hydroxyl (-OH) group. The presence of two singlets at $\delta 0.69$ and 1.01 suggest the existence of two CH₃ attached to the quaternary carbons. The multiplet peaks at $\delta 2.33$ indicates that two methylene protons that are adjacent to the carbon attached to the hydroxyl group. A signal at $\delta 3.55$ presents a proton attached to a hydroxyl (OH) group bearing carbon. This a characteristic proton of steroids (The overlapping triplets at $\delta 5.37$ are assigned to the olefinic proton (a proton attached to a C=C double bond). The FTIR and 1 HNMR spectral patterns suggest that compound AMB3 is most likely a cholest-5-en-3-ol (B-sterol)²⁶. The MHz, CDCl₃) spectrum of ¹³CNMR (101 compound ABM3 (Appendix 11) showed the presence of twenty-nine carbons that include six methyl, eleven methylenes, nine methine and three quaternary carbon signals. The carbon peaks at $\delta 140.75$ and 121.73 could be assigned to the olefinic carbons (C=C). The peak at δ 71.82 indicates a carbon atom bearing hydroxyl (-OH) group. The peaks at $\delta 11.85$ and 19.40 could be assigned to angular methyl carbons (Table No.5). The DEPT-135 spectrum (Appendix 12) compound AMB3 indicates the peaks for the CH₃, and CH while peaks that appear in the $\delta 20-40$ region

indicate the presence of eleven CH₂ groups. Three signals that do not appear in the DEPT-135 spectrum confirm the presence of three quaternary carbons in compound AMB3. Thus, based on the above FTIR, ¹HNMR, ¹³CNMR and Dept-135 spectral data and comparison with previous literature reports^{26,29-31}, the structure of compound ABM3 was suggested to be β -sitosterol (Figure No.4). Isolation of this compound has not been reported from *Aloe calidophila* plant. The ¹³C-NMR and ¹H-NMR and spectra data of this compound ABM3 (β -sitosterol) from *Aloe calidophila* root extract are reported below (Table No.4).

Antibacterial activity tests result

The antibacterial activity of the crude extracts from root of A. calidophila and isolated compounds (ABM1, ABM2 and ABM3) were tested at the concentration of 200 and 300µg/ml against four pathogenic bacterial strains: two Gram-positive bacteria (S. aureus and S. pyogenes) and two Gramnegative bacteria (E. coli and P. auruginosa) (Appendix 13). The result revealed that dichloromethane/methanol (1:1 v/v) and methanol crude extract showed the zone of inhibition ranging from 7 to 9mm for gram-positive bacteria and from 6.5 to 8mm for gram-negative bacteria. The dichloromethane/Methanol (1:1v/v) extract showed higher antibacterial activity against Gram-positive bacteria such as S. aureus (8 mm) and S. pyogenes (8.5 mm) than Gram-negative bacteria such as E. coli (7.5mm) and P. auruginosa (7mm) at the concentration of 300 mg/ml. When dichloromethane/Methanol (1:1v/v) extract is compared with methanol extract against all selected pathogenic strain, methanol extract showed relativelv higher inhibition zone at both concentrations (200 and 300mg/ml) (Table No.5). The isolated compounds showed inhibition zones ranging from 8 to 13mm for gram-positive and from 7.5 to 11.5mm for gram-negative bacteria. The isolated compounds showed significant activities specially in compound ABM3 (Aloesaponarin I) at 300µg/ml against S. aureus, S. pyogen, E. coli and P. auruginosa with inhibition zone of 12, 13, 11.5 and 10.5mm, respectively, compared to positive

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control (Ampicillin) (13.5, 14.5, 13 and 12.5mm, respectively). The data showed that the isolated compounds (ABM1, ABM2 and ABM3) possess relatively higher bacterial zones of inhibition as compared to the crude extracts when tested against the abovementioned bacterial strains (Table No.6). The isolated pure compound showed promising activities against S. pyogenes and S. aureus with ABM1 (Aloesaponarin I) and ABM2 (Oleic acid) showing relatively the best antibacterial activity against S. pyogenes when compared to the rest of one compound ABM3 (β-sitosterol) (Table No.6). Comparatively, compound ABM1 (Aloesaponarin I) has revealed better antibacterial activities against the four strains. These finding is consistent with previous literature²⁷. As described in literatures, this compound occurs in the roots of many Aloe species including A. also saponaria and showed antimicrobial activities²⁴. Compound ABM3 (Aloesaponarin I), was reported to be a promising compound to be used as a lead for development of antimalarial drugs as it showed high antimalarial chloroquine-resistant activity against strain plasmodium parasite³². Finally, the observed antibacterial activities from root of A. calidophila crude extracts and the isolated compounds could support the traditional use of the plant for the treatment of different human illnesses including bacterial infection and malaria reported in literature¹⁸. Thus, further tests are recommended on large number of bacterial strains to determine the potentials of crude extracts and compounds isolated from Aloe calidophila in order to get compounds that can be used in the discovery of new antibacterial agents.

Table No.1: Phytochemical screening test results of the dichloromethane/methanol (1:1) and methanol extracts of Aloe calidophila root

	Secondary		Extracts		
S.No	metabolites	Types of test	Dichloromethane/met hanol (1:1)	Methanol	
1	Alkaloids	Mayer's reaction	+	+	
2	Anthraquinones	Borntragers	+	+	
3	Flavonoids	Alkaline reagent	+	+	
4	Glycosides	Keller-kiliani	+	+	
5	Phenols	Ferric chloride	+	+	
6	Saponins	Froath Foam	+	+	
7	Steroids	Liebermann-Burchardt	_	-	
8	Tannins	Alcholic ferric chloride	-	-	
9	Terpenoids	Liebermann-Burchardt	+	+	

(+): presence of constituent; (-): absence of constituent Table No.2: Comparison of the ¹HNMR (400 MHz, DMSO) and ¹³C NMR (101 MHz, DMSO) data of the compound ABM1 (Aloesaponarin I) and reported data

Position	NMR spectral data of compound ABM1 (ppm)			Literature reported spectral data of Aloesaponarin I ^{24,25}		
	¹³ C-NMR data	¹ H-NMR data	DEPT-135 data	¹³ C-NMR data	¹ H-NMR data	
1	161.89			161.5		
2	112.45	7.30 (dd)	CH (112.45)	112.0	7.31 (dd)	
3	136.57	7.61 (d)	CH (136.57)	136.8	7.62 (t)	
4	118.82	7.0 (t)	CH (118.82)	118.1	7.77 (dd)	
5	130.07	7.72 (s)	CH(124.91)	130.82	7.80 (s)	
6	159.33			159.52		
7	122.89			122.2		
8	117.13			117.5		
9	189.65			189.6		
10	182.21			182.3		
11	132.77			132.8		
12	124.91			124.5		
13	137.17			138.8		
14	141.46			142.91		
-COO	167.64			167.96		
-OCH ₃	52.99	3.88 (s)	CH ₃ (52.99)	52.79	4.06 (s)	
-CH ₃	20.36	2.75 (s)	CH ₃ (20.36)	20.37	2.95 (s)	
1-OH	12.78	12.78 (s)			12.91 (s)	
6-OH	11.78	11.78 (s)			10.45 (s)	

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Position	NMR Spectra data of compound ABM2 (ppm)			Literature Spectra data ²⁷⁻²⁹			
	¹³ C-NMR data	¹ H-NMR data	DEPT-135 data	¹³ C-NMR data	¹ H-NMR data		
1	179.42	-	-	179.7	-		
2	33.94	2.37 (t, 2H)	33.95 (CH ₂)	34.0	2.36 (t, 2H)		
3	24.67	1.65 (m, 2H)	24.68 (CH ₂)	24.68	1.63 (m, 2H)		
4	29.14	1.27 (m, 2H)	29.15 (CH ₂)	29.14	1.27 (m, 2H)		
5	29.08	1.27 (m, 2H)	29.08 (CH ₂)	29.07	1.27 (m, 2H)		
6	29.40	1.27 (m, 2H)	29.42 (CH ₂)	29.4	1.27 (m, 2H)		
7	29.59	1.33 (m, 2H)	29.59 (CH ₂)	29.6	1.32 (m, 2H)		
8	27.15	2.07 (m, 2H)	27.05 (CH ₂)	27.16	2.02 (m, 2H)		
9	127.90	5.38 (d, 1H)	128.07 (CH)	129.7	5.4 (d, 1H)		
10	130.03	5.38 (d, H)	130.03 (CH)	130.03	5.4 (d, 1H)		
11	27.21	2.07 (m, 2H)	27.21 (CH ₂)	27.22	2.02 (m, 2H)		
12	29.70	1.27 (m, 2H)	29.70 (CH ₂)	29.70	1.27 (m, 2H)		
13	29.36	1.27 (m, 2H)	29.35 (CH ₂)	29.33	1.27 (m, 2H)		
14	29.55	1.27 (m, 2H)	29.53 (CH ₂)	29.53	1.27 (m, 2H)		
15	29.24	1.27 (m, 2H)	29.24 (CH ₂)	29.25	1.27 (m, 2H)		
16	31.93	1.27 (m, 2H)	31.93 (CH ₂)	31.94	1.27 (m, 2H)		
17	22.68	1.27 (m, 2H)	22.70 (CH ₂)	22.68	1.27 (m, 2H)		
18	14.12	0.90 (t,3H)	$14.08 (CH_3)$	14.13	0.88, t		

Table No.3: The ¹HNMR (400 MHz, CDCl₃), ¹³CNMR (101 MHz, CDCl₃) and DEPT-135 spectral data of the compound ABM2 and literature reported data of oleic acid²⁷⁻²⁹

Table No.4: Comparison of the ¹H-NMR (400 MHz, CDCl₃) and ¹³CNMR (101 MHz, CDCl3) data of the compound ABM3 (β-sitosterol) and reported data

Position	NMR Spectra data of compound ABM3 (ppm)		Literature NMR spectra data of β- sitosterol ^{27,30-32}		
	¹³ CNMR data	¹ HNMR data	¹³ CNMR data	¹ H-NMR data	
1	37.25		37.3		
2	31.92		31.9		
3	71.82	3.55 (m)	71.8	3.52 (m)	
4	42.29		42.3		
5	140.75		140.8		
6	121.73	5.37 (s)	121.7	5.36 (br s)	
7	31.71		31.7		
8	32.23		32.0		
9	50.14		50.1		
10	37.00		36.6		
11	21.09		21.1		
12	39.79		39.8		
13	42.39		42.4		
14	56.12		56.7		
15	24.3		24.3		
16	28.61		28.6		

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17	56.07		56.1	
18	11.85	0.69(s)	11,9	0.69 (s)
19	19.40	1.01(s)	19.4	1.01 (s)
20	36.15		36.2	
21	14.12	0.90 (dd,6.8)	18.8	0.92 (s)
22	34.84		34.0	
23	26.14		26.1	
24	45.84		45.8	
25	29.15		29.2	
26	19.82	0.84 (d,6.65)	19.8	0.814 (d, 6.5)
27	19.03	0.86 (d,6.65)	19.0	0.833 (d, 6.5)
28	23.06		23.1	
29	11.98	0.87 (t)	12.0	0.845 (t. 7.5)

 Table No.5: Antibacterial inhibition zones (mm) of crude extracts and isolated compounds of Aloe calidophila

	Tested samples		Bacteria species and zone of inhibitions (mm)			
S.No		Sample code	Gram-positive		Gram-negative	
		and Concentration	S. aureus ATCC2592 3	S. pyogen ATCC196 15	E. coli ATCC259 22	P.aeruginosa ATCC27853
1	Dichloromethane/Met	A1=200ug/ml	7	7	6.5	6.5
1	hanol Extract	A2= 300ug/ml	8	8.5	7.5	7
2	Methanol Extract	B1=200ug/ml	7.5	8	7	7
		B2= 300ug/ml	9	9	8	8
2	Compound ABM ₃	C1= 200ug/ml	8	9	8	7.5
5		C2= 300ug/ml	9.5	10	9	9
4	Compound ABM ₂	D1= 200ug/ml	9	10	9	8
		D2= 300ug/ml	10	11	10	9.5
5	Compound ABM ₁	E1= 200ug/ml	10.5	11	10	9
		E2=300ug/ml	12	13	11.5	10.5
6	Ampicillin (positive control)		13.5	14.5	13	12.5

Key: +ve= positive, S. aureus= Staphylococcus aureus, S. pyogenes=Streptococcus pyogenes, E. coli=Escherichia coli, P.aeruginosa=Pseudomonas aeruginosa





Figure No.1. The leaf (a) and root part (b and c) of *Aloe calidophila* (Photo taken by Abinet M on March 19, (2023



Figure No.2: The proposed structure of compound ABM1 (Aloesaponarin I)

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Figure No.4: The proposed structure of compound ABM3 (β-sitosterol)

CONCLUSION

phytochemical screening tests of The the dichloromethane/Methanol (1:1 v/v) and methanol crude extracts of roots of A. calidophila revealed presence of alkaloids. anthraquinones, the flavonoids, phenols, terpenoids, steroids and saponins. The chromatographic separation of dichloromethane/Methanol (1:1 v/v) root extract of calidophila result in isolation of three Α. compounds namely aloesaponarin I (ABM1), oleic acid (ABM2) and β-sitosterol (ABM3). Their structures were determined based on spectral (IR and NMR) data and comparison of the data with literature reports. These compounds were isolated for the first time from A. calidophila. The crude extracts and isolated compounds were subjected to in vitro test to evaluate their antibacterial activities using disc diffusion method against four bacterial strains specifically (E. coli ATCC25922, S. aureus ATCC25923, P. aeruginosa ATCC27853 and S. pyogenes ATCC19615). The results revealed that isolated compounds possess promising the antibacterial activities with aloesaponarin I (compound AMB1) with relatively the highest activity against all the bacterial strains used in the study. Therefore, the further in vitro antibacterial

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activity tests are recommended using several bacterial strains to discover lead compounds from the roots *A. calidophila* in search of new antibacterial agents.

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CONFLICT OF INTEREST

We declare that we have no conflict of interest.

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